

CHROM. 12,176

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### **Rapid separation of guanine-derived compounds using thin-layer chromatography on polyethyleneimine-impregnated cellulose**

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(Received April 17th, 1979)

The separation of guanine-derived compounds is of importance in the field of cyclic nucleotide research. Several methods have been developed to separate the nucleotides and nucleosides using column (anion-exchange and cation-exchange resins), thin-layer and adsorption chromatography<sup>1–6</sup>. Thin-layer chromatographic methods have concentrated upon polyethyleneimine (PEI)-impregnated cellulose. Bohme and Shultz<sup>1</sup> have reviewed these PEI chromatographic procedures utilizing both inorganic and organic solvent systems. The present paper investigates a PEI-cellulose thin-layer chromatographic separation procedure utilizing a single organic buffering system. The system possesses the advantage of rapidity, highly reproducible separation characteristics, and buffer volatility.

## EXPERIMENTAL

### *Chromatographic procedures*

PEI-impregnated cellulose MN 300 (0.1 mm thick) was utilized for all separation procedures. The plates were stored at 4° in a desiccated environment prior to use. The plates were cut from pre-formed 20 × 20 cm plastic backed sheets to a size of approximately 10 × 5 cm for chromatographic separations. The standard guanine-derived compounds were dissolved in water or dilute acid and placed on thin-layer plates as discrete spots measuring 3 mm in diameter. The spots were rapidly air-dried using a hot-air blower. The plates were subsequently placed in a rectangular, buffer-saturated chromatography chamber and developed in an ascending fashion until the solvent front had moved 8–9 cm. The plates were removed from the buffer chamber, air-dried and the standard compounds localized under ultraviolet light illumination (254 nm).

### *Materials*

Triethylamine was purchased from Aldrich (Milwaukee, Wisc., U.S.A.) and redistilled prior to use. Guanosine 5'-triphosphate (GTP), 5'-diphosphate (GDP),

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5'-monophosphate (GMP), guanosine 3',5'-monophosphate (cyclic GMP) and guanosine were all purchased from Sigma (St. Louis, Mo., U.S.A.). Pre-coated plastic backed polygram CEL 300 PEI plates (0.1 mm PEI-cellulose MN 300) were purchased from Brinkman Instruments (Westbury, N.Y.).

#### Buffer preparation

The chromatographic plates were developed using 0.5 M triethylammonium bicarbonate (TEAB) buffer, pH 7.6. This buffer, made from a weak organic base, was routinely prepared every two weeks. A 140-ml volume of distilled triethylamine was diluted to 750 ml with triple deionized, distilled water. A stream of pure carbon dioxide was bubbled through the solution until a pH of 7.6 was reached. The TEAB solution was subsequently brought to 1 l (*i.e.*, 1 M) with water and stored in the dark at room temperature until use. For actual chromatographic separation, the 1 M TEAB stock was diluted to 0.5 M using triple deionized, distilled water. The pH of the buffer was always ascertained to be 7.6 before use.

#### RESULTS AND DISCUSSION

A representative chromatogram of assorted guanine-derived compounds may be seen in Fig. 1. The separation of the nucleotides and nucleosides is based upon pH-dependent charge interaction with the basic PEI<sup>2</sup>. The more highly anionic nucleotides (GTP and GDP) show very little migration while the lesser charged compounds traverse a greater distance. The separations in 0.5 M TEAB developed chromatograms were quite similar to that seen in chromatograms developed by multiple successive buffer systems which utilize acetic acid, water and two different molarities of LiCl<sup>1</sup>.

The reproducibility of the separation may be seen in Table I. The  $R_F$  values have been determined from multiple independent chromatographic separations. The mean values were computed from samples which were either run singly or together

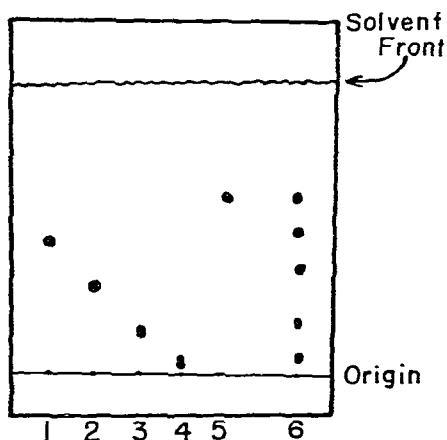


Fig. 1. Representative PEI separation of guanine-derived compounds 1 = cyclic GMP; 2 = GMP; 3 = GDP; 4 = GTP; 5 = guanosine; 6 = mixture of compounds 1-5. Running time with TEAB buffer, pH 7.6 was 40 min with the solvent front traversing 8 cm.

TABLE I

**R<sub>F</sub> VALUES OF GUANINE-DERIVED COMPOUNDS**

The values presented represent the mean  $\pm$  S.E.M. for multiple determinations. The individual determinations included samples run singly in one lane and multiple standards run in the same lane.

<i>Compound</i>	<i>R<sub>F</sub> (<math>\pm</math>S.E.M.)</i>
GTP	0.05 $\pm$ .005 ( <i>n</i> = 18)
GDP	0.18 $\pm$ .01 ( <i>n</i> = 6)
GMP	0.33 $\pm$ .01 ( <i>n</i> = 17)
Cyclic GMP	0.48 $\pm$ .01 ( <i>n</i> = 26)
Guanosine	0.60 $\pm$ .01 ( <i>n</i> = 7)

in a single chromatographic lane. The standard errors of the mean (*i.e.*, precision) were very small, indicative of the reproducibility of the separation. The average time for separation of the above compounds ranged around 40 min for 8 cm migration. Buffer evaporation from the plate following removal from the chromatography tank was extremely rapid due to the volatile nature of TEAB.

Recovery of cyclic GMP from the PEI-cellulose plates can be achieved with 0.15 *M* NaCl. This enables the rapid chromatographic separation to be utilized either directly (with labelled precursors) in a guanylate cyclase assay or as a preliminary step in further purification of cyclic GMP free of contamination. The use of this system adds to the armamentarium of existing thin-layer procedures presently available. It should be emphasized that this single 40-min separation achieves results comparable to the lengthy multiple buffer systems.

## ACKNOWLEDGEMENTS

We would like to acknowledge the excellent technical assistance of Diane Dunlap and manuscript preparation of Catherine Johns.

This work was supported by a NCI grant No. CA 21927 (A.L.M.).

## REFERENCES

- 1 E. Bohme and G. Schultz, *Methods Enzymol.*, 38 (1974) 27.
- 2 K. Randerath, *Thin-Layer Chromatography*, Academic Press, New York, 1964, p. 185.
- 3 N. Krishnan and G. Krishna, *Anal. Biochem.*, 70 (1976) 18.
- 4 G. Schultz, J. G. Hardman, K. Schultz, J. W. Davis and E. W. Sutherland, *Proc. Natl. Acad. Sci. U.S.A.*, 70 (1973) 1721.
- 5 J. Tomasz, *J. Chromatogr.*, 169 (1979) 466.
- 6 N. D. Goldberg and M. K. Haddox, *Methods Enzymol.*, 38 (1974) 73.